

High-Resolution Field Asymmetric Waveform Ion Mobility Spectrometry Using New Planar Geometry Analyzers

Alexandre A. Shvartsburg, Fumin Li, Keqi Tang, and Richard D. Smith*

Biological Sciences Division, Pacific Northwest National Laboratory, MS K8-98, 3335 Q Avenue, Richland, Washington 99352

Field asymmetric waveform ion mobility spectrometry (FAIMS) has emerged as a powerful tool of broad utility for separation and characterization of gas-phase ions, especially in conjunction with mass spectrometry (MS). In FAIMS, ions are filtered by the dependence of mobility on electric field while being carried by gas flow through the analytical gap between two electrodes of either planar (p-) or cylindrical (c-) geometry. Most FAIMS/MS systems employ c-FAIMS because of its ease of coupling to MS, yet the merits of the two geometries have not been compared in detail. Here, a priori simulations reveal that reducing the FAIMS curvature always improves resolution at equal sensitivity. In particular, the resolving power of p-FAIMS exceeds that of c-FAIMS, typically by a factor of 2–4 depending on the ion species and carrier gas. We have constructed a new planar FAIMS incorporating a curtain plate interface for effective operation with an ESI ion source and joined to an MS using an ion funnel interface with a novel slit aperture. The resolution increases up to 4-fold over existing c-FAIMS, even though the analysis is ~2 times faster. This allows separation of species not feasible in previous FAIMS studies, e.g., protonated leucine and isoleucine or new bradykinin isomers. The improvement for protein conformers (of ubiquitin) is less significant, possibly because of multiple unresolved geometries.

The need to analyze samples of ever-growing complexity, primarily in biological applications, has been driving the continued innovation in separation technologies.^{1,2} Separations in condensed media such as liquid chromatography (LC),³ capillary electrophoresis,⁴ strong cation exchange (SCX),⁵ and gel electrophoresis⁶ have been developed extensively and are well-known in the field. These techniques, and particularly their combinations, are exceptionally powerful, with the peak capacity of two-dimensional

methods^{7–9} reaching $\sim 10^3$ – 10^4 . However, separations in condensed phases where molecular diffusion is slow inevitably require a long time (many hours for SCX/LC or 2-D gel^{7–9}), limiting the throughput to several samples per day.

The desire for greater throughput has intensified the interest in gas-phase separations based on ion mobility, which are extremely fast (typically taking milliseconds to seconds) thanks to the high speed of ion motion in gases. The technique of ion mobility spectrometry (IMS) established in 1970s sorts ions by mobility (K) at low electric fields.^{10–13} The other approach of field asymmetric waveform IMS (FAIMS) was known since 1980s^{14–18} but became topical in the past few years.^{19–48} Most recently,

- (1) Smith, R. D.; Shen, Y. F.; Tang, K. Q. *Acc. Chem. Res.* **2004**, *37*, 269.
- (2) Evans, C. R.; Jorgenson, J. W. *Anal. Bioanal. Chem.* **2004**, *378*, 1952.
- (3) Shen, Y.; Zhao, R.; Berger, S. J.; Anderson, G. A.; Rodriguez, N.; Smith, R. D. *Anal. Chem.* **2002**, *74*, 4235.
- (4) Shen, Y.; Xiang, F.; Veenstra, T. D.; Fung, E. N.; Smith, R. D. *Anal. Chem.* **1999**, *71*, 5348.
- (5) Washburn, M. P.; Wolters, D.; Yates, J. R. *Nat. Biotechnol.* **2001**, *19*, 242.
- (6) Shevchenko, A.; Wilm, M.; Vorm, O.; Mann, M. *Anal. Chem.* **1996**, *68*, 850.

- (7) Wolters, D. A.; Washburn, M. P.; Yates, J. R. *Anal. Chem.* **2001**, *73*, 5683.
- (8) Shen, Y. F.; Jacobs, J. M.; Camp, D. G.; Fang, R. H.; Moore, R. J.; Smith, R. D.; Xiao, W. Z.; Davis, R. W.; Tompkins, R. G. *Anal. Chem.* **2004**, *76*, 1134.
- (9) Gygi, S. P.; Corthals, G. L.; Zhang, Y.; Rochon, Y.; Aebersold, R. *Proc. Natl. Acad. Sci. U.S.A.* **2000**, *97*, 9390.
- (10) Hill, H. H.; Siems, W. F.; St. Louis, R. H.; McMinn, D. G. *Anal. Chem.* **1990**, *62*, A1201.
- (11) Bowers, M. T.; Kemper, P. R.; von Helden, G.; van Koppen, P. A. M. *Science* **1993**, *260*, 1446.
- (12) Shvartsburg, A. A.; Hudgins, R. R.; Dugourd, P.; Jarrold, M. F. *Chem. Soc. Rev.* **2001**, *30*, 26.
- (13) Hoaglund-Hyzer, C. S.; Lee, Y. J.; Counterman, A. E.; Clemmer, D. E. *Anal. Chem.* **2002**, *74*, 992.
- (14) Gorshkov, M. P. USSR Inventor's Certificate No. 966583, 1982.
- (15) Buryakov, I. A.; Krylov, E. V.; Soldatov, V. P. USSR Inventor's Certificate No. 1412447, 1988.
- (16) Buryakov, I. A.; Krylov, E. V.; Soldatov, V. P. USSR Inventor's Certificate No. 1485808, 1989.
- (17) Buryakov, I. A.; Krylov, E. V.; Nazarov, E. G.; Rasulev, U. K. *Int. J. Mass Spectrom. Ion Processes* **1993**, *128*, 143.
- (18) Carnahan, B. L.; Tarassov, A. S. U.S. Patent 5420424, 1995.
- (19) Purves, R. W.; Guevremont, R.; Day, S.; Pipich, C. W.; Matyjaszczyk, M. S. *Rev. Sci. Instrum.* **1998**, *69*, 4094.
- (20) Guevremont, R.; Purves, R. W. *Rev. Sci. Instrum.* **1999**, *70*, 1370.
- (21) Guevremont, R.; Purves, R. W.; Barnett, D. A. U.S. Patent 6,713,758, 2003.
- (22) Ells, B.; Froese, K.; Hruday, S. E.; Purves, R. W.; Guevremont, R.; Barnett, D. A. *Rapid Commun. Mass Spectrom.* **2000**, *14*, 1538.
- (23) Barnett, D. A.; Ells, B.; Guevremont, R.; Purves, R. W.; Viehland, L. A. *J. Am. Soc. Mass Spectrom.* **2000**, *11*, 1125.
- (24) Handy, R.; Barnett, D. A.; Purves, R. W.; Horlick, G.; Guevremont, R. *J. Anal. At. Spectrom.* **2000**, *15*, 907.
- (25) Ells, B.; Barnett, D. A.; Purves, R. W.; Guevremont, R. *Anal. Chem.* **2000**, *72*, 4555.
- (26) Barnett, D. A.; Purves, R. W.; Ells, B.; Guevremont, R. *J. Mass Spectrom.* **2000**, *35*, 976.
- (27) Ells, B.; Barnett, D. A.; Purves, R. W.; Guevremont, R. *J. Environ. Monit.* **2000**, *2*, 393.
- (28) Guevremont, R.; Barnett, D. A.; Purves, R. W.; Vandermeij, J. *Anal. Chem.* **2000**, *72*, 4577.
- (29) Gabryelski, W.; Wu, F.; Froese, K. L. *Anal. Chem.* **2003**, *75*, 2478.

FAIMS was coupled to LC⁴⁹ or IMS, creating first 2-D gas-phase separations.⁵⁰ In FAIMS, ions are distinguished by the difference between mobilities at high and low electric fields (E), which in general differ as ion mobilities in gases depend on the field:

$$K(E) = K(0)(1 + a(E/N)^2 + b(E/N)^4 + c(E/N)^6 + \dots) \quad (1)$$

where N is the gas number density. In a typical FAIMS regime ($E/N < 100$ Td), the terms up to $b(E/N)^4$ generally represent $K(E)$ with sufficient accuracy. Depending on the relative values of a and b coefficients, $K(E)$ may increase, decrease, or first increase and then decrease, with corresponding ions classified as types A, C, and B, respectively. These designations are not absolute but depend on the buffer gas: ions exhibiting A-type behavior in one gas may become C-type in another.²³ For example,²³ the protonated heptadecanoic acid is A-type in CO₂ or N₂O but C-type in O₂ or N₂. Most FAIMS studies have used air or N₂ gases, but gas mixtures such as He/CO₂, N₂/CO₂, and especially He/N₂ are attracting increasing interest. The resolution and sensitivity of FAIMS using binary and ternary mixtures is often better than that with any individual component,^{25–27,33,48} because of non-Blanc behavior of ion mobilities at high electric fields.⁴⁸ Addition of volatile vapors to a homomolecular buffer gas produces a similar effect.⁴¹

FAIMS is implemented by flowing a gas stream containing ions through the space between two electrodes (the “analytical gap”) carrying a periodic asymmetric waveform $V_D(t)$ plus a dc compensation voltage (CV).^{19,20} The waveform induces ion oscillation in the gap, and the displacements during positive and negative segments of $V_D(t)$ slightly differ because of unequal mobilities at high and low E . This would result in ions migrating toward one of the electrodes (depending on the ion type) and neutralizing

on impact. For a particular species, a certain CV may produce an offsetting drift, allowing ions to remain in equilibrium inside the gap.^{19,20} Scanning the CV reveals the spectrum of ionic mixture. The earliest FAIMS devices used a planar analytical gap confined between two parallel electrodes (p-FAIMS).^{14–17} An alternative configuration with ions separated in the annular space between two coaxial cylinders (c-FAIMS) was demonstrated subsequently.^{16,18–20} A geometry comprising two concentric spheres was also proposed.²¹ While the spherical design had not been tested in experiment, a “dome” geometry (c-FAIMS plus a hemispherical section) has become widely used.^{22–35} The introduction of micromachined FAIMS “chips” gave rise to miniaturized p-FAIMS devices.^{38–43}

The electric field in a curved analytical gap is inhomogeneous and, thus, may focus ions to the gap median. However, $V_D(t)$ of either polarity focuses ions of one type (A or C) only but removes those of the other type from the gap.^{19,20} Hence, acquiring a full CV spectrum using c-FAIMS requires scanning CV with both $V_D(t)$ polarities, which halves the instrumental duty cycle. Further, the focusing strength depends on the slope of $K(E)$ curve over the relevant E range: ions with a near-flat $K(E)$ (often belonging to type B) focus poorly and are depleted in c-FAIMS. This phenomenon distorts the CV spectra measured using c-FAIMS, discriminating against ions with lower absolute CV.⁵¹ Homogeneous electric fields cannot focus ions, so the above adverse behaviors are not encountered in p-FAIMS.

Both planar and cylindrical (“dome” and “side-by-side”^{21,52}) geometries have been implemented in commercial FAIMS systems, the DMS^{41,43} (Sionex, Bedford, MA) and Selectra^{33,34} (Ionalytics, Ottawa, Canada), with the number and diversity of applications growing swiftly. These span the fields such as detection of drugs, explosives, and chemical warfare agents,^{31,35,36,40} environmental monitoring,^{22,24,25,27,29,39} bacterial typing,⁴² product quality assurance,³² natural resource management,³⁰ and biomedical research.³³ However, the utility of FAIMS has been restricted by resolving power^{46,47} (R) of ~ 10 – 20 (compared to R of ~ 150 in the best IMS designs).^{53–55} This has limited the separation of even simple mixtures, e.g., H³⁴SO₄[–] and ³⁵ClO₄[–] isobars^{24,27} or three phthalic acid isomers.²⁶ While those species can still be distinguished via buffer gas modifications,^{26,27} such changes have to be made for each individual case by trial and error, and the overall peak capacity may not increase. The resolution may also be improved by reducing the analytical gap width (g) or the waveform frequency (ω_c),⁴⁷ superposing a low-frequency ripple⁵⁶ onto $V_D(t)$, or varying the hemispherical gap width in the “dome” design,³⁵ but all of these approaches also diminish sensitivity.^{35,56} Importantly, the resolution cannot be raised beyond a certain limit (depending on CV and the $V_D(t)$ amplitude known as the dispersion voltage, V_{\max}) even as ion transmission through FAIMS approaches zero.⁵⁶

- (30) Gabryelski, W.; Froese, K. L. *Anal. Chem.* **2003**, *75*, 4612.
 (31) McCooney, M. A.; Mester, Z.; Ells, B.; Barnett, D. A.; Purves, R. W.; Guevremont, R. *Anal. Chem.* **2003**, *74*, 3071.
 (32) McCooney, M.; Ding, L.; Gardner, G. J.; Fraser, C. A.; Lam, J.; Sturgeon, R. E.; Mester, Z. *Anal. Chem.* **2003**, *75*, 2538.
 (33) Cui, M.; Ding, L.; Mester, Z. *Anal. Chem.* **2003**, *75*, 5847.
 (34) Guevremont, R. *J. Chromatogr., A* **2004**, *1058*, 3.
 (35) Guevremont, R.; Thekkadath, G.; Hilton, C. K. *J. Am. Soc. Mass Spectrom.* **2005**, *16*, 948.
 (36) Buryakov, I. A. *Talanta* **2003**, *61*, 369.
 (37) Buryakov, I. A. *J. Chromatogr., B* **2004**, *800*, 75.
 (38) Miller, R. A.; Nazarov, E. G.; Eiceman, G. A.; King, A. T. *Sens. Actuators, A* **2001**, *91*, 301.
 (39) Eiceman, G. A.; Tarassov, A.; Funk, P. A.; Hughs, S. E.; Nazarov, E. G.; Miller, R. A. *J. Sep. Sci.* **2003**, *26*, 585.
 (40) Krylov, E.; Nazarov, E. G.; Miller, R. A.; Tadjikov, B.; Eiceman, G. A. *J. Phys. Chem. A* **2002**, *106*, 5437.
 (41) Eiceman, G. A.; Krylov, E. V.; Krylova, N. S.; Nazarov, E. G.; Miller, R. A. *Anal. Chem.* **2004**, *76*, 4937.
 (42) Schmidt, H.; Tadjimukhamedov, F.; Mohrenz, I. V.; Smith, G. B.; Eiceman, G. A. *Anal. Chem.* **2004**, *76*, 5208.
 (43) Veasey, C. A.; Thomas C. L. P. *Analyst* **2004**, *129*, 198.
 (44) Krylov, E. V. *Tech. Phys.* **1999**, *44*, 113.
 (45) Krylov, E. V. *Int. J. Mass Spectrom.* **2003**, *225*, 39.
 (46) Shvartsburg, A. A.; Tang, K.; Smith, R. D. *J. Am. Soc. Mass Spectrom.* **2004**, *15*, 1487.
 (47) Shvartsburg, A. A.; Tang, K.; Smith, R. D. *J. Am. Soc. Mass Spectrom.* **2005**, *16*, 2.
 (48) Shvartsburg, A. A.; Tang, K.; Smith, R. D. *Anal. Chem.* **2004**, *76*, 7366.
 (49) Venne, K.; Bonneil, E.; Eng, K.; Thibault, P. *Anal. Chem.* **2005**, *77*, 2176.
 (50) Tang, K.; Li, F.; Shvartsburg, A. A.; Strittmatter, E. F.; Smith, R. D. *Anal. Chem.* **2005**, *77*, 6381.

- (51) Barnett, D. A.; Guevremont, R.; Purves, R. W. *Appl. Spectrosc.* **1999**, *53*, 1367.
 (52) Guevremont, R.; Ding, L.; Ells, B.; Barnett, D. A.; Purves, R. W. *J. Am. Soc. Mass Spectrom.* **2001**, *12*, 1320.
 (53) Asbury, G. R.; Hill, H. H. *J. Microcolumn Sep.* **2000**, *12*, 172.
 (54) Srebalus, C. A.; Li, J.; Marshall, W. S.; Clemmer, D. E. *Anal. Chem.* **1999**, *71*, 3918.
 (55) Dugourd, Ph.; Hudgins, R. R.; Clemmer, D. E.; Jarrold, M. F. *Rev. Sci. Instrum.* **1997**, *68*, 1122.
 (56) Shvartsburg, A. A.; Tang, K.; Smith, R. D. *J. Am. Soc. Mass Spectrom.* **2005**, *16*, 1447.

The remaining instrumental parameter is the gap curvature, which (for c-FAIMS) depends on the internal (r_{in}) and external (r_{out}) electrode radii. Increasing r_{in} (at constant $g = 2$ mm) from 4 to 12 mm raises R several times,⁵⁷ because weaker ion focusing at lower curvature narrows the CV range of ions that may pass the gap. Extrapolating that trend to $r_{\text{in}} \rightarrow \infty$, the p-FAIMS with no ion focusing was surmised to provide the highest R , but this was not substantiated or quantified in either theory or experiment. An experimental and modeling investigation of p- and c-FAIMS has shown the latter to provide a higher transmission for ions with strong focusing properties (i.e., high $|CV|$) and lower transmission for other ions.⁴⁵ However, the compared devices were dissimilar in several key aspects in addition to their curvature. First, p- and c-FAIMS used $\{g = 0.5$ mm; $w_c = 1.3$ MHz $\}$ and $\{g = 2$ mm; $w_c = 200$ –250 kHz $\}$, respectively, and performance is quite sensitive to those parameters.⁴⁷ Second, the gap length (L) was 15 mm for p- but 90 mm for c-FAIMS, and the ion residence times (t_{res}) differed by a factor of >100 : ~ 2 ms versus ~ 360 ms, respectively, which unduly favored the p-FAIMS in a sensitivity comparison.^{46,47} While the inevitable tradeoff between resolution and ion transmission means that sensitivities of different designs should be studied for a fixed R , the resolutions achievable for p- and c-FAIMS have not been compared previously. The model⁴⁵ has also neglected the space-charge effects and anisotropy of ion diffusion that are important for a realistic description of FAIMS performance⁴⁶ and made other approximations needed for a closed-form mathematical solution.

The operational regimes of p- and c-FAIMS matched in ref 45 are typical for existing devices (e.g., DMS and Selectra): for p-FAIMS, g is 3–4 times smaller (0.5 vs 1.5–2 mm), w_c is ~ 2 –10 times higher (1–2 MHz vs 80–750 kHz), and t_{res} are shorter by 2 orders of magnitude (~ 1 –5 vs ~ 100 –500 ms). However, these drastic differences reflect a historical legacy not related to inherent properties of the two geometries.

While FAIMS and IMS may work as stand-alone analyzers with a Faraday cup detector,^{14–16,19,20,36} much interest involves their coupling to mass spectrometry (MS). The hemispherical section of “dome” FAIMS that collects ions separated in the cylindrical part provides a natural MS interface, resulting in a broad acceptance of that design.^{22–35} In a planar analytical gap, ions freely spread laterally to the gas flow and only a minor fraction enters a small orifice leading to the MS. This has compelled virtually all FAIMS/MS systems to adopt c-FAIMS, regardless of its intrinsic merits versus those of p-FAIMS.

Here we employ first-principles calculations to compare the performance of planar and cylindrical FAIMS a priori, keeping other factors equal. The modeling has revealed a fundamental advantage of p-FAIMS in terms of the resolution–sensitivity balance, in addition to the duty cycle gain and lack of discrimination discussed above. Guided by those simulations, we have constructed a new p-FAIMS with resolving power several times that of any known FAIMS. This device has been coupled to an MS using a novel “slit–aperture” interface, enabling practical high-resolution FAIMS/MS analyses. The improved peak capacity allowed separating ionic species previously indistinguishable by FAIMS (or IMS).

THEORETICAL FOUNDATION

Computational Treatment. The FAIMS model used herein has been described and extensively validated by measurements.^{46,47,56} The overall concept follows SIMION:⁵⁸ a representative ensemble of ions is propagated through the device by trajectory integration. The statistics of those passed is aggregated, with ion transmission given by the ratio of the number of exiting and initial trajectories. Evaluation of this ratio as a function of CV produces the peak profile and yields the full width at half-maximum (fwhm) and R , defined as (peak CV)/(fwhm).

The underlying physics incorporates ion diffusion (accounting for the high-field and anisotropic components), Coulomb repulsion of ions within the ensemble,⁴⁶ and a nonuniform profile of gas flow velocity across the analytical gap.⁵⁶ This capability was used to probe the dependence of FAIMS response on various design and operational parameters, including g , L , the $V_D(t)$ profile, w_c , and the identity and flow velocity of buffer gas.^{46,47,56} Those assessments were restricted to c-FAIMS with “median radius” $r_{\text{cen}} = (r_{\text{in}} + r_{\text{out}})/2 = 8$ mm (adopted in most analyzers including Selectra), but the model would work for any geometry as long as $E(t)$ and gas flow are defined at any point. Present simulations are for the bisinusoidal waveform^{46,47,56}

$$V_D(t) = [f \sin(w_c t) + \sin(2w_c t - \pi/2)] V_{\text{max}} / (f + 1) \quad (2)$$

with $f = 2$ as used in Selectra, DMS, and the new p-FAIMS reported here. This value of f is optimum for FAIMS performance.⁴⁷ The FAIMS response substantially depends^{46,47,56} on the ion current intensity, I . Here we model the extremes of (i) low I where the Coulomb repulsion is negligible and (ii) saturation where the FAIMS space-charge capacity is filled. The results turn out similar; thus, the conclusions would hold in any real situation. While all calculations below are for homomolecular buffers, they could be extended to heteromolecular media via a formalism treating non-Blanc effects.⁴⁸

Simulations as a Function of FAIMS Analyzer Curvature and Experimental Validation. Since the present computational model was previously corroborated for $r_{\text{cen}} = 8$ mm only,^{46,47,56} first we test its performance as a function of gap curvature using the data for bromochloroacetate (BCA) anion ($m/z = -173$ Da) in N_2 carrier gas.⁵⁷ In practice, increasing r_{cen} from 5 to 9 mm and further to 13 mm (at fixed $g = 2$ mm) narrows the CV spectral peaks at any V_{max} in the $-(1.80$ – $3.96)$ kV range studied,⁵⁷ as shown for minimum and maximum values (Figure 1a–f). This behavior is due to a weaker ion focusing at lower curvature, as described above. However, at low $|V_{\text{max}}|$ the peaks cease to narrow above certain r_{in} , e.g., at $V_{\text{max}} = -1.80$ kV, once a fwhm of 0.2 V is reached at $r_{\text{in}} \sim 8$ mm (Figure 1, parts c and e). This happens because focusing (always weak at low $|V_{\text{max}}|$) becomes negligible at low curvature. Further, increasing $|V_{\text{max}}|$ (at any curvature) has not significantly improved R ; while CVs increase, peaks widen nearly in proportion because of stronger ion focusing at higher $|V_{\text{max}}|$.⁵⁷

We have simulated the evolution of CV spectra for BCA[−] as a function of V_{max} and r_{in} (for experimental⁵⁷ $w_c = 750$ kHz, $T = 298$ K, $a = 7.984 \times 10^{-6}$ Td^{−2}, $b = -3.049 \times 10^{-10}$ Td^{−4}; assuming⁵⁷ $t_{\text{res}} = 0.2$ s and $K(0) = 1.7$ cm²/(V·s) at 298 K), Figure 1a–f. As

(57) Guevremont, R.; Purves, R. W. *J. Am. Soc. Mass Spectrom.* **2005**, *16*, 349.

(58) Dahl, D. A. *Int. J. Mass Spectrom.* **2000**, *200*, 3.

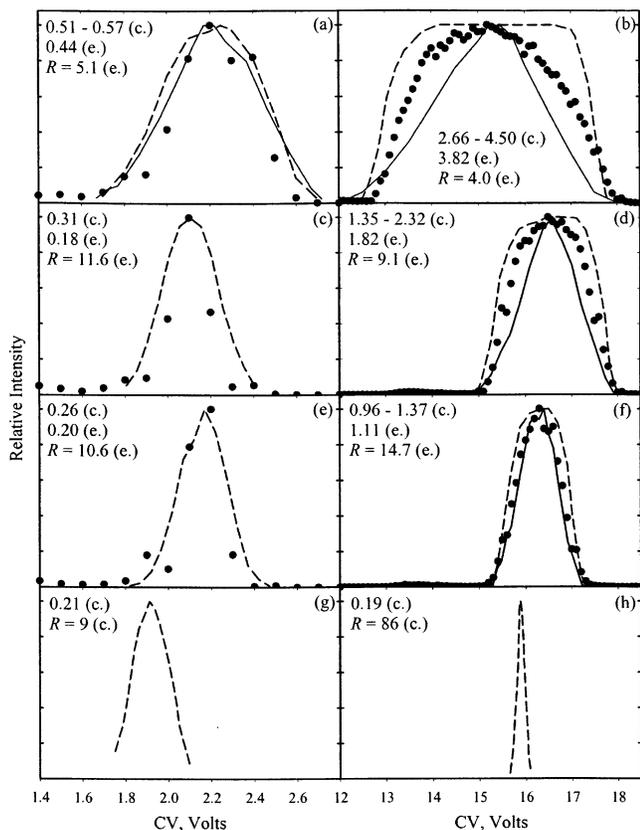


Figure 1. CV peaks for BCA^- at $V_{\text{max}} = -1.80$ kV (left column) and -3.96 kV (right column), for c-FAIMS geometries with $r_{\text{cen}} = 5$ mm (a, b), 9 mm (c, d), and 13 mm (e, f), and p-FAIMS (g, h); $g = 2$ mm for all. Shown are measurements (circles) and present simulations (lines) in the limits of zero (dashed) and saturated (solid) ion current. In (c), (e), (g), and (h), the solid and dashed lines coincide within the computational accuracy. The calculated (c) and experimental (e) values of fwhm (V) and the resolving power are listed in each panel.

usual, features at saturated I are narrower than those in the limit of $I \rightarrow 0$, because Coulomb repulsion preferentially removes from the analytical gap the ions with CVs near the edges of the transmission band that would otherwise pass FAIMS.⁴⁶ At high $V_{\text{max}} = -3.96$ kV, the measured peaks lie between the two theoretical limits. At lowest $V_{\text{max}} = -1.8$ kV, the calculated peaks appear slightly broader than the measured ones (in Figure 1, parts c and e), but the latter are clearly narrowed by having just two–three points across the peak. Overall, the agreement with all observed dependences of peak profiles (and hence R) on both V_{max} and r_{in} is very good.

We have then extended the model to planar FAIMS geometry (Figure 1, parts g and h). When a peak has ceased to sharpen with increasing r_{cen} because of negligible ion focusing at low curvature, it obviously would not narrow further in a p-FAIMS. This is the case at the lowest $V_{\text{max}} = -1.8$ kV, where fwhm = 0.2 V (Figure 1g) as in c-FAIMS with $r_{\text{cen}} = 9$ –13 mm (Figure 1, parts c and e). To the contrary, at high V_{max} (-3.96 kV) the computed peak in p-FAIMS (Figure 1h) is ~ 5 times narrower than that in any c-FAIMS considered previously (Figure 1f) and ~ 10 times narrower than that in the Selectra device (Figure 1d). This is not surprising: absence of ion focusing in planar FAIMS means a constant peak width (here 0.2 V) regardless of V_{max} ; thus, the resolution advantage of p-FAIMS grows with increasing $|V_{\text{max}}|$. The

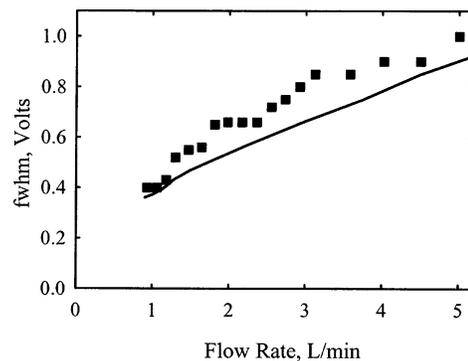


Figure 2. Peak width for toluene ions filtered by micromachined p-FAIMS as a function of gas flow rate: measurements (squares) and present calculation (line).

most striking result is the potential for reaching $R \sim 100$ (Figure 1h), which would bring the FAIMS resolving power close to that of IMS.

One critical difference between p- and c-FAIMS is in the dependence of peak widths on the ion residence time. In c-FAIMS, filtering out ions with CVs outside of a finite range allowing equilibrium in the gap requires a certain t_{res} (~ 50 ms in Selectra), and further FAIMS action does not improve resolution.⁴⁷ In p-FAIMS, there is a single CV permitting ion equilibrium, and ions with even a very small CV difference will eventually be eliminated given sufficient time. Hence, longer separation times would increase R (in principle) indefinitely. To test the ability of present model to quantify that effect, we have simulated the toluene peaks in micromachined p-FAIMS³⁸ with $w_c = 2$ MHz and $V_{\text{max}} = 1.1$ kV (Figure 2). Knowing the gap dimensions^{38,61} of $L = 15$ mm, $S = 10$ mm (span), and $g = 0.5$ mm, the gas flow rate (1–5 L/min) may be converted into linear velocity and further into t_{res} (0.9–4.5 ms). The evolution of features over that time scale was modeled assuming $\{K(0) = 2.3 \text{ cm}^2/(\text{V}\cdot\text{s})$ at 298 K, $a = 1.28 \times 10^{-5} \text{ Td}^{-2}$, $b = -8.2 \times 10^{-10} \text{ Td}^{-4}\}$, with $K(0)$ derived from IMS⁶² and $\{a, b\}$ obtained from FAIMS data.⁴³ Excellent agreement between predicted and measured peak profiles (Figure 2) supports the applicability of the present model to p-FAIMS over a range of separation times.

Relative Intrinsic Merits of Planar and Cylindrical FAIMS. With the simulation validated for FAIMS analyzers of any (and no) curvature, we proceed to compare the response of c- and p-FAIMS with all other factors equal. Benchmarking will be made for the exemplary system of deprotonated leucine anion in air, for which the performance of c-FAIMS (with $r_{\text{cen}} = 8$ mm) was extensively modeled,^{46,47,56} in agreement with experiments. As previously, we assume conditions of ($V_{\text{max}} = -3.3$ kV, $w_c = 210$ kHz, $L = 30$ mm, $t_{\text{res}} = 0.2$ s, $T = 298$ K) and use $\{K(0) = 2.18 \text{ cm}^2/(\text{V}\cdot\text{s})$ at 298 K, $a = 5.43 \times 10^{-6} \text{ Td}^{-2}$, $b = -1.85 \times 10^{-10} \text{ Td}^{-4}\}$ ^{46,47} derived from FAIMS measurements.^{59,60} Similarly to the behavior in Figure 1, increasing r_{in} (at constant $g = 2$ mm, in the $I \rightarrow 0$ limit) raises R from 10 at $r_{\text{cen}} = 8$ mm to ~ 45 at $r_{\text{cen}} = 73$ mm (Figure 3a). The

(59) Barnett, D. A.; Ells, B.; Guevremont, R.; Purves, R. W. *J. Am. Soc. Mass Spectrom.* **1999**, *10*, 1279.

(60) Guevremont, R.; Barnett, D. A.; Purves, R. W.; Viehland, L. A. *J. Chem. Phys.* **2001**, *114*, 10270.

(61) Miller, R. A.; Eiceman, G. A.; Nazarov, E. G.; King, A. T. *Sens. Actuators, B* **2000**, *67*, 300.

(62) Lubman, D. M. *Anal. Chem.* **1984**, *56*, 1298.

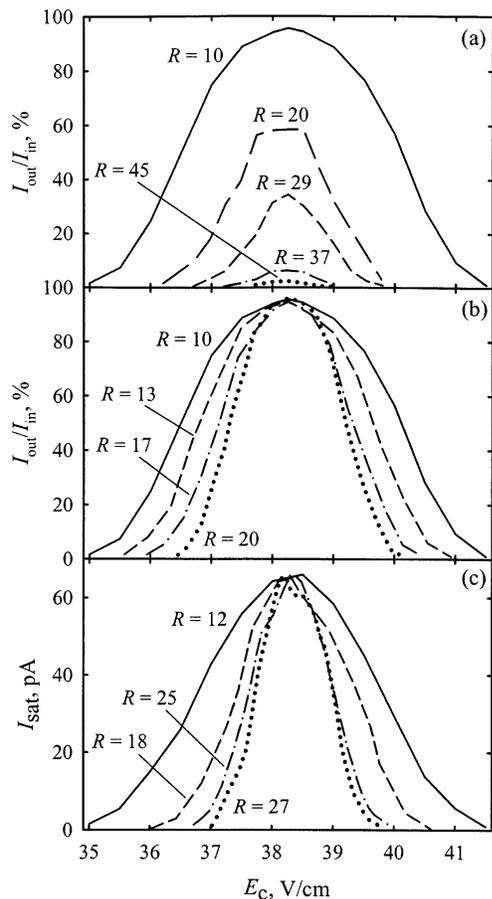


Figure 3. Simulated CV peaks for (leucine - H)⁻ as a function of analytical gap curvature, for the limits of zero ion current (a), and saturated current (c). From outside to inside, curves are for $r_{cen} = 8, 13, 18, 38, \text{ and } 73$ mm in (a); $8, 18, 38, \text{ and } \infty$ (p-FAIMS) in (b), and $8, 19, 49, \text{ and } \infty$ in (c). The gap width is fixed at $g = 2$ mm (a) or adjusted to keep constant sensitivity (b, c), with respective g of 2, 2.7, 3.5, and 8.5 mm (b) and 2, 2.4, 2.7, and 2.7 mm (c). To account for a variable g , CV is expressed via compensation field, $E_c = (CV)/g$. The computed values of resolving power are given in all cases.

magnitude of improvement (a factor of 4.5) is less than ~ 10 for BCA^- (Figure 1, part h vs part d): the ion focusing for (leucine - H)⁻ is weaker because of (i) lower $|V_{max}| = 3.3$ kV versus 3.96 kV for BCA^- and (ii) lower CV at equal V_{max} due to a less steep $K(E)$ curve with the value of a equal to $\sim 70\%$ of that for BCA^- . A weaker ion focusing with increasing r_{cen} also leads to greater losses, reducing ion transmission from 96% at $R = 10$ to 2% at $R \sim 45$ (Figure 3a). Further curvature reduction to p-FAIMS results in yet lower transmission and has not been modeled.

Hence the question is how to exploit the resolution advantage of (more) planar FAIMS without sacrificing sensitivity? This may be achieved by widening the gap while reducing its curvature. For example, the ion transmission of $\sim 95\%$ (at $I \rightarrow 0$) may be retained by increasing g from 2 to ~ 8.5 mm as r_{cen} is raised from 8 mm to infinity (Figure 3b). Then R improves less dramatically, doubling from 10 for standard c-FAIMS to 20 for p-FAIMS. This also occurs in the other limit of saturated I (Figure 3c), with R at equal ion intensity increasing from 12 to ~ 27 . Again, the effect is greater in cases of higher CV (that correspond to stronger ion focusing in c-FAIMS). Therefore, the transition to planar geometry could improve R at any ion intensity 2-fold or more with no

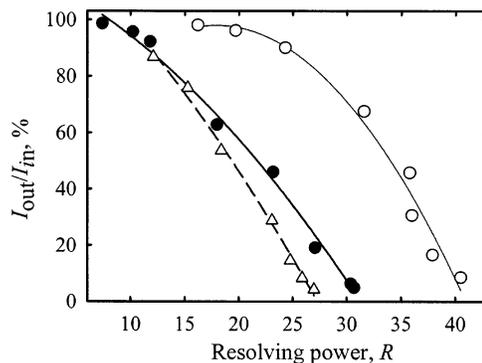


Figure 4. Calculated resolution-sensitivity tradeoff (at $I \rightarrow 0$) for (leucine - H)⁻ in p-FAIMS (empty circles) and c-FAIMS with $R_{cen} = 8$ mm (filled circles) (ref 56), both obtained by varying the gap width. Triangles are for the tradeoff obtained in c-FAIMS by scaling a 60 Hz "ripple" harmonic superposed on the asymmetric waveform (ref 56). Lines are quadratic regressions through the data for each curve.

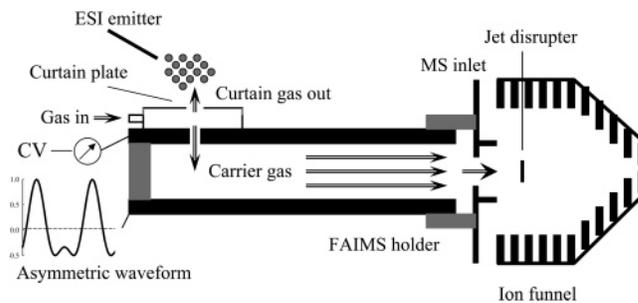


Figure 5. Scheme of the new planar FAIMS analyzer coupled to an MS using an ion funnel interface.

sensitivity impairment. Accordingly, the curve summarizing the tradeoff between FAIMS resolution and sensitivity (Figure 4) moves to the right when the gap curvature is reduced. The greatest gain is at highest sensitivity, moderating at lower sensitivity (and higher R). This happens because higher FAIMS resolution is achieved by constraining the gap, meaning a lower curvature of c-FAIMS. The advantage of p-FAIMS is yet greater when compared versus c-FAIMS with R controlled using ripple⁵⁶ (Figure 4).

IMPLEMENTATION OF A NEW HIGH-RESOLUTION PLANAR FAIMS

While, by theory, p-FAIMS allows a higher resolution, it is not achievable using existing designs with insufficient ion residence times. In fact, the resolution of DMS p-FAIMS was generally worse than that of Selectra c-FAIMS, as the difference of t_{res} by a factor of ~ 100 outweighed the intrinsic resolution advantage. Also, the DMS chips do not provide for the desolvation of ions generated by ESI and thus are less suitable for ESI-MS coupling.

Hence we have constructed a new p-FAIMS analyzer permitting much longer separations (Figure 5). In this device, the analytical gap is formed by two polished stainless steel plates secured inside a plastic (PEEK) enclosure and precisely aligned by ceramic spacers. The gap dimensions are $g = 2$ mm (same as in most c-FAIMS systems including Selectra), $L \sim 50$ mm, and $S = 20$ mm. Thus, the cross-sectional area is 40 versus 113 mm² in Selectra, meaning a proportionately shorter t_{res} at equal gas flow rate, e.g., ~ 0.1 s versus the ~ 0.2 s typical in Selectra. Still, that exceeds the t_{res} in previous p-FAIMS by ~ 20 –100 times, which should improve resolution.

The new p-FAIMS also features an effective ESI/FAIMS interface (Figure 5). Ions enter the gap through a curtain plate chamber resembling that in Selectra,⁵⁷ with curtain plate and sampling apertures of 2.5 and 1.5 mm, respectively. The carrier gas enters through a side opening in the chamber and splits into two streams: the major (~70%) flows out of the curtain plate orifice and desolvates incoming ions while the minor carries ions into the sampling orifice and through the gap (Figure 5). To enable FAIMS/MS analyses, the device was installed in a custom PEEK holder secured to the inlet of time-of-flight (TOF) mass spectrometer. The FAIMS exit opposes the sampling aperture, leaving a break of ~0.5 mm for electrical insulation. The gas is supplied to the device by a unit that formulates gas mixtures of up to three components (each predried by hygroscopic filters) and controls the flow rate in the 0.5–5 L/min range. Here we employed N₂ and He/N₂ mixtures with 10–50% He (v) at total flow rates of 2–3 L/min. Then the flow through the gap is ~0.5–0.8 L/min, approximately matching the MS inlet conductance. The ESI emitter is mounted a few millimeters away from the inlet on an X–Y translation stage for fine position adjustment (Figure 5). Samples are infused to the emitter (at 0.4 μ L/min) through a metal union using a microsyringe driven by a pump (kd Scientific, Holliston, MA).

The bisinusoidal asymmetric waveform (eq 2) with $w_c = 750$ kHz and $f = 2$ is produced by a power supply similar to that in Selectra, that adds the 750 and 1500 kHz harmonics (with 2:1 amplitude ratio) output by resonating LC circuits.⁶³ The CV is generated by a programmable dc power supply. The $V_D(t)$ and CV are applied to opposite electrodes, simplifying the arrangement in Selectra where they are coapplied to the internal electrode. The V_{max} value is adjustable from 2.3 to 4 kV; here we use 3.9 kV. The whole FAIMS unit is biased at 190 V, the curtain plate is at 1 kV, and the ESI emitter is at ~3 kV (referenced to ground). Spectra are obtained by scanning the desired CV range with a speed of 0.5–5 V/min. All instrumental parameters are controlled by custom software resident on a dedicated PC. Since there is no ion focusing, the $V_D(t)$ polarity is immaterial and (unlike for c-FAIMS) need not be switched depending on the ion type.

The TOF MS is an LC/MSD TOF (Agilent Technologies, Palo Alto, CA) with the capillary skimmer cone at the front end replaced by electrodynamic ion funnel that significantly raises the ion utilization efficiency at API/MS interfaces.^{64–66} The present funnel has 100 round electrodes with i.d.'s reducing from 25 mm at the "mouth" to 2 mm at the exit to MS, and features a jet disrupter⁶⁶ that improves sensitivity. The funnel chamber is evacuated by a mechanical pump (pumping on the inlet skimmer chamber in the original TOF). The capillary inlet normally leading into the ion funnel is replaced by an aperture cut through a 0.2 mm steel sheet (Figure 5). We have tested two apertures—a circle of 0.43 mm diameter and a noncontiguous "slit" made of 11 circular 0.13 mm holes disposed uniformly along a 4 mm segment. These apertures

(63) Krylov, E. V. *Instrum. Exp. Tech.* **1997**, *40*, 628.

(64) Kim, T.; Tolmachev, A. V.; Harkewicz, R.; Prior, D. C.; Anderson, G. A.; Udseth, H. R.; Smith, R. D.; Bailey, T. H.; Rakov, S.; Futrell, J. H. *Anal. Chem.* **2000**, *72*, 2247.

(65) Tang, K.; Shvartsburg, A. A.; Lee, H. N.; Prior, D. C.; Buschbach, M. A.; Li, F.; Tolmachev, A. V.; Anderson, G. A.; Smith, R. D. *Anal. Chem.* **2005**, *77*, 3330.

(66) Tang, K.; Tolmachev, A. V.; Nikolaev, E.; Zhang, R.; Belov, M. E.; Udseth, H. R.; Smith, R. D. *Anal. Chem.* **2002**, *74*, 5431.

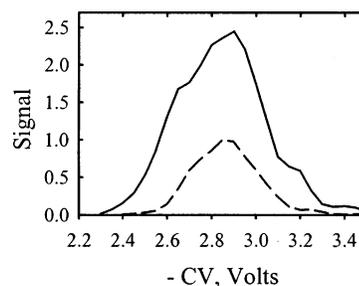


Figure 6. CV spectra of H⁺reserpine measured using p-FAIMS with the circular (dash) and "slit" (solid line) MS inlet apertures.

have the same cross-sectional area of 0.145 mm², resulting in equal funnel chamber pressure of 2 torr. The rationale for slit aperture is to increase the spatial overlap with the ribbon-shaped ion beam exiting p-FAIMS. The funnel focuses and guides broad or divergent ion beams in a pseudopotential well created by combination of dc and rf voltages applied to the electrodes.^{64–66} Here the voltages were 190 V on the MS aperture, 187–40 V dc along the funnel, and 175 V on the jet disrupter; the peak rf amplitude was 45 V (at 560 kHz). All voltages above are for positive ions and should be reversed for negative ion analyses.

For benchmarking purposes, p-FAIMS was replaced by the Selectra c-FAIMS with no other changes to the system. The exit orifice of Selectra is circular; hence, the circular MS inlet aperture was used. The $V_D(t)$ waveform and gap width in Selectra and present p-FAIMS are identical, allowing a direct performance comparison ($V_{max} = 4$ kV or -4 kV was chosen for A- and C-type ions, respectively). The resolution of Selectra may be tuned by longitudinal translation of the internal electrode that adjusts the axial gap width in hemispherical region (g_H) from 1.7 to 2.7 mm in 0.1 mm increments.³⁵ Increasing g_H above $g = 2$ mm improves the FAIMS resolution but lowers sensitivity.³⁵ Here we used the maximum practical^{35,50} $g_H = 2.3$ –2.5 mm to provide a stringent benchmark for resolution of new p-FAIMS; the comparison with Selectra operated at more typical $g_H \sim 2.1$ mm⁵⁰ would favor p-FAIMS even more.

INCREASED RESOLUTION OF NEW PLANAR FAIMS

First, we compared the merits of two MS inlet apertures described above. The ion signal with the "slit" was consistently >250% of that with the circular aperture (Figure 6). While this clearly shows the advantage of elongated apertures at the p-FAIMS/MS interface, our present "slit" design can be refined further, and its optimization is in progress.

The separation performance of new p-FAIMS has been characterized using several representative analytes: 10 μ M reserpine, 50 μ M 1:1 leucine/isoleucine, 10 μ M bradykinin, and 50 μ M bovine ubiquitin, all obtained from Sigma Aldrich. For infusion into ESI, those chemicals were dissolved in 50:49:1 methanol/deionized water/acetic acid.

Reserpine is a common standard for MS system evaluation. With N₂ buffer, the CV spectrum of H⁺reserpine ($m/z = 609$ Da) comprises a dominant feature at -3 V and two lesser ones at -4 V and ~ -4.5 V (Figure 6). To compare the resolution of two FAIMS geometries, we have chosen the major peak. As He content grows from 0% to 50%, the CV in c-FAIMS moves from -3 V to

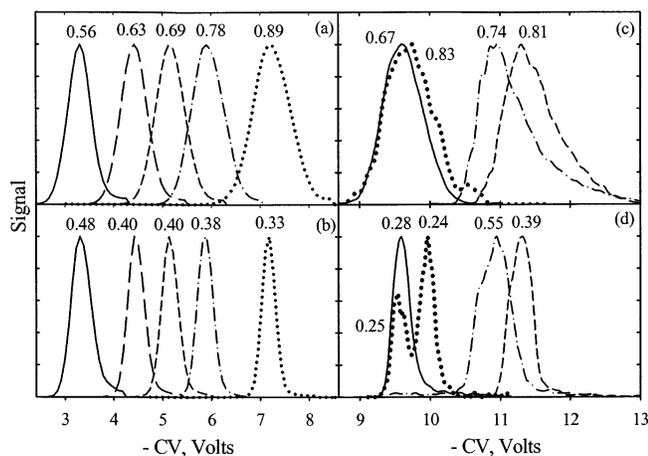


Figure 7. CV spectra of H^+ reserpine (a, b) and H^+ leucine/ H^+ isoleucine (c, d) acquired using Selectra (a, c) [with $V_{\max} = -4$ kV (a) and 4 kV (c)] and p-FAIMS (b, d), in N_2 (solid line) and He/N_2 with a He fraction of 20% (long dash), 30% (short dash), 40% (dash-dot), and 50% (dotted). For clarity, we plot only the major peak for reserpine (at CV = -3 V in Figure 6). Spectra in (b, d) were aligned with those in (a, c) to match the CVs at peak apexes. Measured fwhm values are given for all features.

-7 V while the width increases from 0.6 to 0.9 V (Figure 7a). The peak broadening at higher $|CV|$ is typical of c-FAIMS; stronger ion focusing at greater CV widens the CV transmission window. This does not happen in p-FAIMS; in fact the features narrow from 0.5 V in N_2 to 0.3 V at 50% He (Figure 7b) because at higher He concentration $K(0)$ increases, effectively constraining the gap. Overall, the resolution advantage of p-FAIMS is by a factor of 1.2–2.7, which is close to the above theoretical prediction.

IMS and FAIMS (typically in conjunction with MS) are often employed to characterize isomeric mixtures. For example, a prototypical bioanalytical problem is distinguishing leucine and isoleucine. These amino acids were previously separated by c-FAIMS as negative deprotonated ions⁵⁹ but not as positive protonated ions ($m/z = 132$ Da) for which the forms of $K(E)$ (and thus CVs) differ less.⁶⁰ We also could not separate H^+ isoleucine from H^+ leucine using Selectra (Figure 7c). In N_2 , there is a single peak of ~ 0.7 V width at CV = -9.6 V, consistent with previous data.^{59,60} As discussed above, ions in He/N_2 tend to exhibit non-Blanc effects that often improve separation.^{33,48} Here, CV shifts with increasing He fraction to more negative values up to -11.3 V at $\sim 30\%$ He, then the trend reverses with -9.7 V reached at 50% He (Figure 7c). Such a maximum of absolute CV (evidencing non-Blanc behavior) is similar to that for many other species⁴⁸ in He/N_2 , but the peak width remains at ~ 0.7 – 0.8 V and there is no separation improvement. In p-FAIMS, the peaks are consistently sharper (0.25–0.55 V wide) at any He fraction (Figure 7d). This suffices to clearly separate the two isomers at 50% He, though not at lower He concentrations. The increase of R is from 12 to 15 in c-FAIMS (Figure 7c) to 20–42 in p-FAIMS (Figure 7d), i.e., by a factor of 2.5 on average: again close to theoretical predictions.

One promising FAIMS application is the analysis of proteolytic digests.^{28,49,50,67} Global proteome digests can consist of 10^5 – 10^6

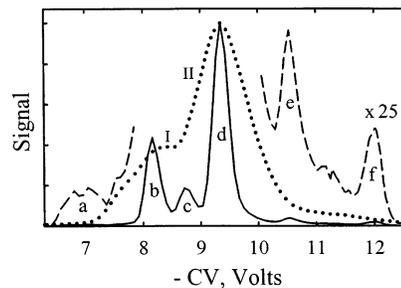


Figure 8. CV spectra of $(H^+)_2$ bradykinin obtained using Selectra with $V_{\max} = -4$ kV (dotted) and p-FAIMS (full spectrum, solid line; smaller features expanded by $\times 25$, dashed line).

different peptides,⁶⁸ and the need for their rapid characterization is the major driver for continued innovation in separations science. The speed of gas-phase ion mobility methods makes them extremely attractive for high-throughput proteomics platforms based on multidimensional separations, such as LC/FAIMS/MS⁴⁹ and FAIMS/IMS/MS.⁵⁰ Ion mobility approaches may also distinguish isomeric/isobaric peptides, including sequence inversions.⁶⁹ Since tryptic peptides cover a limited CV range, a modest FAIMS resolution has particularly constrained its utility in proteomics. A classical model peptide in MS and IMS studies is bradykinin (RPPGFSPFR), which under soft ESI conditions produces mainly $(H^+)_2$ bradykinin⁷⁰ ($m/z = 531$ Da). Two conformers of this ion (minor I and major II at higher $|CV|$) were distinguished using the Selectra,⁷⁰ the result reproduced here (Figure 8). However, combined FAIMS and H/D exchange data expose four different species.⁷⁰ Using p-FAIMS, we resolved six features (a–f) in the CV spectrum for $(H^+)_2$ bradykinin (Figure 8). The isomers b and d match, respectively, I and II, but we discover new isomers a in the ledge of I, c hidden between I and II, and e and f in the extended tail of II (Figure 8). Well-defined features b and d are ~ 0.3 V wide, meaning $R \sim 30$, which exceeds the R of Selectra by a factor of 3–4 (as expected from simulations). The species a, b, c, and d apparently correspond to 4, 3, 2, and 1 distinguished by FAIMS–H/D exchange,⁷⁰ whereas e and f were not reported previously.

Finally, we have applied the new p-FAIMS to characterize macromolecular conformers. Proteins and other flexible macromolecules may adopt a plurality of 3-D structures in solution that are partly conserved by soft ionization mechanisms such as ESI.^{71–73} This allows using IMS and FAIMS to explore protein polymorphism and isomeric transitions (denaturation) in solution depending on the temperature, solvent composition, acidity (ionic strength), and presence of specific chemicals, including as a function of time.^{71–73} The growing interest in misfolded proteins (prions) as the cause of transmissible spongiform encephalopathies, Alzheimer's, and other neurodegenerative diseases has made this research increasingly topical.⁷⁴ All methods for protein

(67) Barnett, D. A.; Ells, B.; Guevremont, R.; Purves, R. W. *J. Am. Soc. Mass Spectrom.* **2002**, *13*, 1282.

(68) Aebersold, R.; Mann, M. *Nature* **2003**, *422*, 198.

(69) Wu, C.; Siems, W. F.; Klasmeier, J.; Hill, H. H. *Anal. Chem.* **2000**, *72*, 391.

(70) Purves, R. W.; Barnett, D. A.; Ells, B.; Guevremont, R. *Rapid Commun. Mass Spectrom.* **2001**, *15*, 1453.

(71) Li, J.; Taraszka, J. A.; Counterman, A. E.; Clemmer, D. E. *Int. J. Mass Spectrom.* **1999**, *185/186/187*, 37.

(72) Purves, R. W.; Barnett, D. A.; Guevremont, R. *Int. J. Mass Spectrom.* **2000**, *197*, 163.

(73) Hudgins, R. R.; Woenckhaus, J.; Jarrold, M. F. *Int. J. Mass Spectrom. Ion Processes* **1997**, *165/166*, 497.

structure elucidation (in either solution or gas phase) face a fundamental challenge: all proteins exist in solution as an ensemble of numerous similar but slightly different geometries. This causes blurring of NMR structures for proteins in solution, a diminished resolution of LC or CE separations, and broadening of IMS peaks that limits R to $\sim 30\text{--}35$, whatever the instrumental performance.^{71,73,75} The resolution of FAIMS should be affected likewise, but the actual limitation could not be determined because of low instrumental R .

Ubiquitin is a common small protein (76 residues, 8565 Da for bovine) often used as a model for biophysical investigations. ESI of acidified ubiquitin solutions generally yields ions of charge states (z) of 6+ to 14+, with most having several isomers distinguished by IMS⁷¹ and FAIMS.^{72,76–78} Here we focus on $z = 10\text{--}13$, where the protein is largely unfolded (by Coulomb repulsion) resulting in simpler CV spectra that allow a direct comparison between c- and p-FAIMS. In agreement with earlier measurements,^{72,78} the data acquired using Selectra show peaks at CV = $-(8\text{--}9)$ V (Figure 9). The peak widths are 0.8–1.0 V for 10+ and 13+ and ~ 1.5 V for 11+ and 12+ that exhibit poorly resolved features,^{72,78} also with fwhm of 0.8–1.0 V as determined by Gaussian peak fitting. Switching to p-FAIMS reduces the fwhm for 10+ and 13+ to 0.4–0.7 V but reveals no new conformers (Figure 9). Four and two features are partly resolved for 11+ and 12+, respectively, with fwhm of 0.3–0.7 V. These more structured spectra are close to those measured⁷⁷ using Selectra at $V_{\text{max}} = -4.4$ kV, with peaks $\sim 0.4\text{--}0.5$ V wide. Hence the resolution advantage of p-FAIMS for ubiquitin is only $\sim 1.5\text{--}2$ times, i.e., less than that for bradykinin with similar CVs (Figure 8) though comparable to that for leucine/isoleucine (Figure 7 c, d). These data suggest $\sim 0.3\text{--}0.4$ V as the minimum peak width for protein ions set by multiplicity of exact geometries making up a “single” conformer; firm clarification of this issue is pending detailed studies involving other proteins.

CONCLUSION

A comprehensive modeling of FAIMS separations based on first-principles molecular dynamics has revealed a uniform improvement of performance (in terms of resolution/sensitivity tradeoff) with reduction of analytical gap curvature. In particular, planar (p-) FAIMS is superior to cylindrical (c-) FAIMS, always providing a higher resolving power at equal ion utilization efficiency, for any ion current intensity. The resolution advantage increases with increasing absolute CV of an ion, with a gain of $\sim 2\text{--}4$ -fold over commercial c-FAIMS instrumentation for typical analytes and carrier gases. In contrast to c-FAIMS, the resolution of p-FAIMS is highly sensitive to the duration of analysis over any time scale, and given sufficient time, much higher resolution may be achieved. These results are in a quantitative agreement with FAIMS measurements as a function of curvature and ion residence time.

(74) Bernstein, S. L.; Wyttenbach, T.; Baumketner, A.; Shea, J. E.; Bitan, G.; Teplow, D. B.; Bowers, M. T. *J. Am. Chem. Soc.* **2005**, *127*, 2075.

(75) Shvartsburg, A. A.; Li, F.; Tang, K.; Smith, R. D. *Anal. Chem.*, DOI:10.1021/ac060283z.

(76) Purves, R. W.; Barnett, D. A.; Ells, B.; Guevremont, R. *J. Am. Soc. Mass Spectrom.* **2000**, *11*, 738.

(77) Purves, R. W.; Barnett, D. A.; Ells, B.; Guevremont, R. *J. Am. Soc. Mass Spectrom.* **2001**, *11*, 894.

(78) Robinson, E. W.; Williams, E. R. *J. Am. Soc. Mass Spectrom.* **2005**, *16*, 1427.

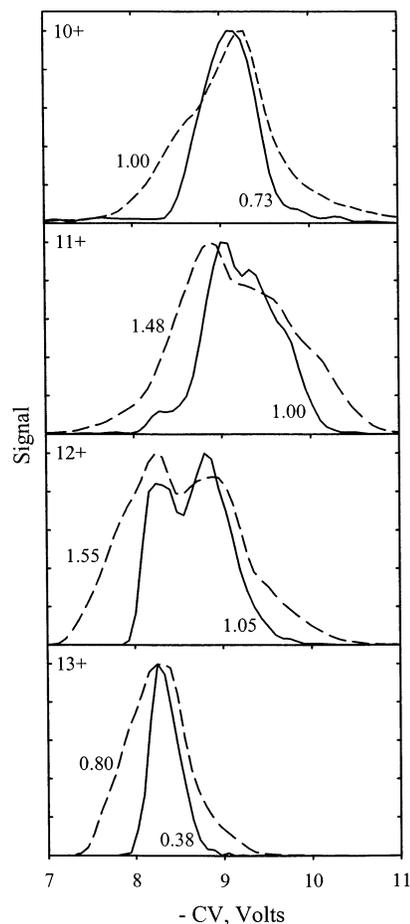


Figure 9. CV spectra for protonated bovine ubiquitin ($z = 10\text{--}13$) obtained using Selectra with $V_{\text{max}} = -4$ kV (dash) and p-FAIMS (solid line). Measured fwhm are indicated.

Guided by simulations, we have developed and demonstrated a planar FAIMS with separation time scales approaching that of the Selectra c-FAIMS (~ 0.1 s), i.e., $\sim 20\text{--}100$ times longer than in existing miniature p-FAIMS devices. As predicted by theory, the resolution of new system exceeds that of c-FAIMS in all cases. The gain ranges from $\sim 20\%$ to a factor of ~ 4 (generally increasing at higher $|CV|$), with peaks narrowing to 0.25 V and the resolving power reaching 40. A proportional increase of peak capacity allows p-FAIMS to deal with significantly more complex analytes. In an isomeric separation, protonated leucine and isoleucine that have a CV difference of $\sim 3.5\%$ could be distinguished by p-FAIMS but not the Selectra. With application to conformational analyses, p-FAIMS has found four new conformers of the bradykinin dication, in addition to the two separated by c-FAIMS. The improvement for proteins (exemplified by ubiquitin) is lesser, likely because the resolution becomes limited by the multiplicity of conformers occupying neighboring minima on the potential energy surface rather than instrumental factors, as previously seen in IMS. Still, p-FAIMS has resolved ubiquitin conformations not found using c-FAIMS under otherwise identical conditions.

Broad use of planar FAIMS has been impeded by the lack of practical ESI and MS interfaces. We have integrated a curtain plate ESI interface into p-FAIMS and developed a new elongated slit-ion funnel MS interface that captures a ribbonlike ion beam delivered by p-FAIMS, converts it into a circular shape, and

effectively conveys it to MS analyses. The "slit" aperture substantially increases the sensitivity of FAIMS/MS, and the ongoing optimization of aperture geometry is expected to yield further improvements.

In conclusion, the new planar design enables high-resolution FAIMS with the peak capacity approaching that of modern IMS. Since FAIMS is intrinsically more orthogonal to MS than IMS, this development makes FAIMS/MS a promising tool for many applications. Unlike in IMS, no a priori means to deduce ionic structures from FAIMS data currently exist. However, FAIMS pre-separation greatly augments the structural elucidation power of IMS,^{50,75} and the work to couple planar FAIMS to IMS/MS is in progress.

ACKNOWLEDGMENT

The authors thank Dr. Roger Guevremont and Dr. Randy Purves (ThermoElectron, Ottawa, Canada) for numerous insightful discussions of FAIMS technology and providing the original experimental data. This work was supported by the NIH National Center for Research Resources (RR 18522). PNNL is a multiprogram national laboratory operated by the Battelle Memorial Institute for the U.S. Department of Energy through Contract DE-AC05-76RLO1830.

Received for review November 14, 2005. Accepted April 4, 2006.

AC052020V